Phycicoccus endophyticus sp. nov., an endophytic actinobacterium isolated from *Bruguiera gymnorhiza*

Shao-Wei Liu,¹ Min Xu,² Li Tuo,¹ Xiao-Jun Li,^{1,3} Lin Hu,⁴ Li Chen,⁴ Rong-Feng Li⁵ and Cheng-Hang Sun¹

¹Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing 100050, PR China

²Faculty of Basic Medical Sciences, Guilin Medical University, Guilin 541004, PR China
³College of Laboratory Medical Science, Hebei North University, Zhangjiakou 075000, PR China
⁴Institute of Zoology, Chinese Academy of Sciences, Beijing 100101, PR China
⁵Department of Chemistry, Johns Hopkins University, Baltimore, MD 21218, USA

A novel endophytic actinobacterium, designated strain IP6SC6^T, was isolated from surfacesterilized bark of Bruguiera gymnorhiza collected from Zhanjiang Mangrove Forest National Nature Reserve in Guangdong, China. Cells of strain IP6SC6^T were Gram-stain-positive, aerobic, non-spore-forming, non-motile rods. Strain IP6SC6^T grew at 20-42 °C (optimum, 37 °C), at pH 6.0-9.0 (optimum, pH 7.0) and in the presence of 0-8 % (w/v) NaCl (optimum, 0-2 %). Chemotaxonomic analyses showed that the isolate possessed meso-diaminopimelic acid as the diamino acid of the peptidoglycan, galactose and glucose as whole-cell sugars, and MK-8(H₄) as the predominant menaquinone. The major polar lipids were diphosphatidylglycerol, phosphatidylinositol and an unknown lipid. The major fatty acids were iso-C_{15:0}, anteiso- $C_{15:0}$, anteiso- $C_{17:0}$ and iso- $C_{16:0}$. The G+C content of the genomic DNA was 72.5 mol%. Phylogenetic analyses based on 16S rRNA gene sequences revealed that strain IP6SC6^T belonged to the genus Phycicoccus and shared the highest sequence similarity with *Phycicoccus jejuensis* NRRL B-24460^T (96.97 %). On the basis of phylogenetic analysis and phenotypic and chemotaxonomic characteristics, strain IP6SC6^T represents a novel species of the genus Phycicoccus, for which the name Phycicoccus endophyticus sp. nov. is proposed. The type strain is $IP6SC6^{T}$ (=DSM 100020^T=CGMCC 4.7300^T).

The genus *Phycicoccus*, a member of the family *Intrasporangiaceae*, suborder *Micrococcineae* (Stackebrandt *et al.*, 1997), was created by Lee (2006) and the description was subsequently emended by Zhang *et al.* (2011). At the time of writing, the genus comprises eight species with validly published names: *Phycicoccus jejuensis* (Lee, 2006) as the type species, *P. dokdonensis* (Yoon *et al.*, 2008), *P. aerophilus* (Weon *et al.*, 2008), *P. bigeumensis* (Dastager *et al.*, 2008), *P. cremeus* (Zhang *et al.*, 2011), *P. ginsenosidimutans* (Wang *et al.*, 2011), *P. badiiscoriae* (Lee, 2013) and *P. soli* (Singh *et al.*, 2015). Members of the genus *Phycicoccus* are Gram-stain-positive, aerobic coccoid- or rod-shaped bacteria isolated from diverse environments, such as dried seaweed, air, soil and scoria. The characteristics of the genus *Phycicoccus* include *meso*-diaminopimelic acid as the diagnostic diamino acid in the cell-wall peptidoglycan, MK-8(H₄) as the major menaquinone, and iso- $C_{15:0}$ and iso- $C_{16:0}$ as the predominant fatty acids. The G+C content of the genus *Phycicoccus* is in the range 69.7–74.0 mol% (Lee, 2013).

During an investigation of the culturable actinobacterial diversity associated with endophytic actinomycetes from mangrove plants, strain IP6SC6^T, the first endophytic actinobacterium affiliated with the genus *Phycicoccus*, was isolated from surface-sterilized bark of *Bruguiera gymnor*-*hiza* collected from Zhanjiang Mangrove Forest National Nature Reserve (21° 34′ 14″ N 109° 45′ 21″ E) in Guang-dong, China. Based on polyphasic taxonomic studies, strain IP6SC6^T was distinguished from previously described species of the genus *Phycicoccus* and is considered to represent a novel species. In this paper, the taxonomic position of this strain is reported.

Correspondence Cheng-Hang Sun chenghangsun@hotmail.com or sunchenghang@imb.pumc.edu.cn

The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain $IP6SC6^{T}$ is KT327645.

Two supplementary tables and four supplementary figures are available with the online Supplementary Material.

Treatment of the plant sample was processed as described by Qin *et al.* (2009). After sterilization, the sample was ground to a powder by using a micromill and distributed on International Streptomyces Project (ISP) 2 agar (Shirling & Gottlieb, 1966) supplemented with 1 % (v/v) plant tissue extract. After 4 weeks of incubation at 28 °C, a pure culture was isolated and then subcultured on trypticase soy agar (TSA; BD). The purified isolate was maintained at 4 °C on TSA slants and preserved in aqueous glycerol suspensions (20 %, v/v) at -80 °C.

The extraction of genomic DNA from strain IP6SC6^T and PCR amplification of the 16S rRNA gene were performed as described by Li et al. (2007). The purified PCR products were cloned using the pEASY-T1 Cloning kit (TransGen Biotech) according to the manufacturer's instructions, and sequenced using an ABI PRISM 3730XL DNA Analyser. Levels of 16S rRNA gene sequence similarity between strain IP6SC6^T and related species were determined via the EzTaxon e-server (http://eztaxon-e.ezbiocloud.net/; Kim et al., 2012). Multiple alignments were generated using the CLUSTAL X program (Thompson et al., 1997). Evolutionary distances were calculated using the Kimura two-parameter model (Kimura, 1983). Phylogenetic trees were reconstructed using the neighbour-joining (Saitou & Nei, 1987), maximum-parsimony (Fitch, 1971) and maximum-likelihood (Felsenstein, 1981) methods with MEGA version 6.0 (Tamura et al., 2013). The topologies of the phylogenetic trees were evaluated by using the bootstrap method of Felsenstein (1985) with 1000 repeats.

The nearly full-length 16S rRNA gene sequence (1487 bp) of strain IP6SC6^T was obtained to determine its phylogenetic position. Comparative 16S rRNA gene sequence analyses showed that strain IP6SC6^T was phylogenetically most closely related to members of the family Intrasporangiaceae. Strain IP6SC6^T shared highest 16S rRNA gene sequence similarity with *Phycicoccus jejuensis* NRRL B-24460^T (96.97 %), followed by Tetrasphaera duodecadis ATCC 13347^T (96.90 %), *Phycicoccus badiiscoriae* Sco-B23^T (96.65 %), Tetrasphaera veronensis Ver1^T (96.62 %), Phycicoccus bigeumensis MSL03^T (96.46 %), Phycicoccus dokdonensis DS-8^T (96.45 %), Tetrasphaera japonica ACM 5116^T (96.32 %) and Tetrasphaera elongata $Lp2^{T}$ (96.28 %). Lower sequence similarities (<96.0 %) were found with all other recognized species of the family Intrasporangiaceae. The phylogenetic tree reconstructed using the neighbourjoining algorithm (Fig. 1) showed that strain IP6SC6^T was separate from members of the genus Tetrasphaera and fell into the cluster of the genus Phycicoccus with a relatively high bootstrap value (71 %), forming a separate clade within the family Intrasporangiaceae. This topology was also observed in the trees based on the maximum-likelihood and maximum-parsimony algorithms (Figs S1 and S2, available in the online Supplementary Material).

Studies of the cultural, physiological and biochemical characteristics of strain $IP6SC6^{T}$ were carried out under the same conditions with *P. jejuensis* NRRL B-24460^T.

P. jejuensis NRRL B-24460^T, the closest phylogenetic neighbour of strain $IP6SC6^{T}$ in the genus *Phycicoccus*, was obtained from Agricultural Research Service Culture Collection (Peoria, USA) and used as a reference strain. Cultural characteristics were determined by observing growth of the strain at 28 °C for 1-2 weeks on ISP 2, 3, 4, 5 and 7 agars (Shirling & Gottlieb, 1966), nutrient agar (Waksman, 1961), R2A agar (BD), TSA, yeast-starch agar (Ara & Kudo, 2007) and Bennett's agar (Gordon & Smith, 1955). The ISCC-NBS colour charts (Kelly, 1964) were used to assess colony colour and diffusible pigment. Cell morphology and motility were observed by transmission electron microscopy (JEM-1400; JEOL) after incubation on TSA at 28 °C for 3 days. The Gram-stain test was performed as described by Magee et al. (1975). Growth under anaerobic conditions was determined after incubation in the BBL GasPak Anaerobic System (BD) at 28 °C for 14 days. The temperature range for growth was determined by incubation of the strain on TSA at 4, 15, 20, 25, 28, 30, 37, 42, 45 and 50 °C for 14 days. The pH range for growth was measured in trypticase soy broth (TSB; BD) with various pH values (pH 4.0-12.0, at intervals of 1.0 pH unit) for 14 days. For the pH experiments, the different buffers described by Xu et al. (2005) were used. Salt tolerance was tested in TSA supplemented with 0, 1, 2, 3, 5, 7, 8, 9 or 10 % (w/v) NaCl for 14 days. Catalase activity was determined by bubble production in 3 % (v/v) H_2O_2 . Oxidase activity was assessed by using 1 % (w/v) tetramethyl-p-phenylenediamine (Cappuccino & Sherman, 2002). Hydrolysis of starch, cellulose, gelatin and Tweens 20, 40 and 80, production of H₂S, and milk coagulation and peptonization were examined as described by Gonzalez et al. (1978). Acid production from carbon sources was tested using the API 50CH (bioMérieux) system according to the manufacturer's instructions. Other biochemical characteristics and enzyme activities were tested by using the API 20NE and API ZYM kits (bioMérieux) according to the manufacturer's instructions.

Cells of strain IP6SC6^T were aerobic, Gram-stain-positive, non-spore-forming and non-motile. Colonies grown on TSA plates were circular, smooth, opaque and greyish yellow. Cells of strain IP6SC6^T were rod-shaped (0.5-0.8 µm in diameter and 1.2-2.0 µm in length) on TSA after incubation at 28 °C for 3 days (Fig. S3). Strain IP6SC6^T displayed good growth on TSA, R2A agar, ISP 2 and ISP 3 agars, Bennett's agar and yeast-starch agar, poor growth on ISP 5 and ISP 7 agars, and no growth on ISP 4 agar or nutrient agar. Substrate and aerial mycelia were not observed and no diffusible pigment was produced on any of the media tested. The strain was capable of growth on TSA containing 0-8 % NaCl; it grew at temperatures from 20 to 42 °C and at pH 6.0-9.0. Optimum growth occurred at 37 °C, at pH 7.0 and with 0-2 % (w/v) NaCl. No growth occurred at 15 or 45 °C, at pH 5.0 or 10.0, or in the presence of 9 % (w/v) NaCl. Detailed physiological and biochemical characteristics of strain IP6SC6^T are given in Table 1 and the species description.



Fig. 1. Neighbour-joining phylogenetic tree based on 16S rRNA gene sequences showing the relationships between strain IP6SC6^T and the type strains of related species in the family *Intrasporangiaceae*. *Angustibacter aerolatus* 7402J-48^T was used as an outgroup. Numbers at nodes refer to bootstrap values (based on 1000 replicates; only values >50 % are shown). Bar, 5 nt substitutions per 1000 nt.

For the chemotaxonomic investigations, including polar lipids, menaquinones, diagnostic diamino acids and sugars, biomass of both strain IP6SC6^T and the reference strain P. jejuensis NRRL B-24460^T was obtained after incubation in TSB at 28 °C, 180 r.p.m., for 3 days. The polar lipids were extracted and analysed by two-dimensional TLC on silica gel 60 F₂₅₄ plates (Merck) as described by Minnikin et al. (1984). The solvent systems of the first and the second dimension were chloroform/methanol/water (64: 27: 5, by vol.) and chloroform/methanol/acetic acid/ water (80 : 18 : 12 : 5, by vol.), respectively. Menaguinones were isolated and purified according to the method of Collins et al. (1977), then analysed and identified using an HPLC system coupled to a single quadrupole mass spectrometer (Guo et al., 2015). The diagnostic diamino acids and sugars in whole-cell hydrolysates were identified by TLC as described by Schleifer & Kandler (1972) and Staneck & Roberts (1974), respectively. For the analysis of whole-cell fatty acids, cell mass of strain IP6SC6^T and *P. jejuensis* NRRL B-24460^T were harvested from both TSA and R2A agar plates grown at 28 °C for 3 days, when the bacterial communities reached the late-exponential stage of growth according to the four quadrants streak method. The whole-cell fatty acids were saponified, methylated and extracted according

to the standard protocol described by Sasser (1990), and analysed according to the method described by Tuo *et al.* (2015). For G+C content assessment, genomic DNA of strain IP6SC6^T was prepared according to the method described by Marmur (1961) and the G+C content was determined by reversed-phase HPLC as described by Mesbah *et al.* (1989).

Strain IP6SC6^T contained *meso*-diaminopimelic acid as the diagnostic diamino acid in the cell-wall peptidoglycan, and galactose and glucose as whole cell-wall sugars. The predominant menaquinone was identified as MK-8(H₄) (98.5 %) and the minor component as MK-7(H_4) (1.5 %). The polar lipids were diphosphatidylglycerol, phosphatidylinositol and an unknown lipid (Fig. S4). Although there were some differences in proportions, the profiles of fatty acids in both TSA and R2A agar medium were quite similar. Four major components (>10 % of the total), iso- $C_{15:0}$, anteiso- $C_{15:0}$, anteiso- $C_{17:0}$ and iso- $C_{16:0}$, and some minor components (<10 %), $C_{18:1}\omega_9c$, $C_{16:0}$, iso- $C_{17:0}$, $C_{18:0}$, $C_{16:1}\omega 7c$, iso- $C_{14:0}$, 10-methyl $C_{18:0}$, $C_{14:0}$ and 10-methyl C_{16:0}, were detected in both TSA and R2A agar medium. The comparative cellular fatty acid compositions of strain IP6SC6^T and *P. jejuensis* NRRL B-24460^T in both

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tager et al., 2008); 5, P. dokdonensis DS-8^T (Yoon et al., 2008); 6, P. cremeus V2M29^T (Zhang et al., 2011); 7, P. ginsenosidimutans KCTC 19419^T (Wang et al., 2011); 8, P. aerophilus 5516T-20^T (Weon et al., 2008); 9, P. soli THG-a14^T (Singh et al., 2015). All were positive or weakly positive for catalase, naphthol-AS-BI-phosphohydrolase and β-glucosidase, and assimilation of D-glucose, malate, maltose and N-acetyl-D-glucosamine. All were negative for glucose fermentation, arginine dihydrolase and α -fucosidase, and assimilation of caprate and citrate. +, Positive; -, negative; Strains: 1, IP6SC6^T (data from this study); 2, P. jejuensis NRRL B-24460^T (data from this study, except where indicated); 3, P. badiiscoriae Sco-B23^T (Lee, 2013); 4, P. bigeumensis MSL-03^T (Das-(+), weakly positive; ND, not determined.

Characteristic	1	2	3	4	5	6	7	8	6
Source of isolation	Bark of <i>Bruguiera</i> gymnorhiza	Seaweed*	Scoria	Soil	Soil	Soil	Soil	Air	Soil
Cell morphology Colony colour	Rods Greyish yellow	Cocci* Moderate vellow	Cocci Moderate yellow	Cocci Yellow	Cocci Greyish vellow	Rods Cream	Cocci Greyish vellow	Short rods White	Cocci White
Temperature for growth (°C)	20-42	ycnow 15–37	20-35	20–37	успом 10—36	14–35	успом 10—37	5-37	10-35
pH for growth	6.0-9.0	5.0 - 10.0	5.1-11.1	7.0-12.0	5.0 - 8.5	4.1 - 10.0	5.0 - 10.0	5.0 - 9.0	5.5-8.5
NaCl tolerance for growth (%)	0-8.0	0-8.0	0-1.0	0-5.0	0-5.0	0-7.0	0-5.0	0-7.0	0-4.5
Oxidase	(+)	I	Ι	I	+	I	+	I	+
Hydrolysis of starch	I	I	ND	+	+	I	ND	+	+
Hydrolysis of gelatin	+	+	+	I	+	+	+	+	+
Indole reduction	I	I	1	I	I	I	I	I	+
Nitrate reduction	I	+	I	+	Ι	+	+	Ι	+
Enzyme activity (API ZYM)									
Alkaline phosphatase	I	I	+	+	+	(+)	I	I	+
Esterase (C4)	+	+	1	+	+	I	I	I	+
Esterase lipase (C8)	+	+	(+)	+	+	+	Ι	+	+
Lipase (C14)	I	+	I	I	I	I	I	I	+
Leucine arylamidase	+	+	(+)	+	+	+	I	+	+
Valine arylamidase	(+)	(+)	(+)	Ι	+	+	Ι	Ι	+
Cysteine arylamidase	+	(+)	Ι	Ι	+	Ι	Ι	Ι	+
Trypsin	I	I	(+)	I	Ι	+	I	Ι	1
α -Chymotrypsin	I	+	I	I	I	I	I	+	+
Acid phosphatase	+	+	+	+	+	+	Ι	+	+
α -Galactosidase	I	+	Ι	I	+	+	I	Ι	+
β -Galactosidase	I	+	I	+	+	+	+	+	+
eta-Glucuronidase	I	I	I	+	I	I	I	I	+
α-Glucosidase	+	+	+	I	+	+	I	+	+
N -Acetyl- β -glucosamidase	I	I	Ι	I	I	+	I	I	I
α-Mannosidase	I	I	I	+	(+)	+	I	I	+
Assimilation of (API 20NE):									
D-Arabinose	I	I	I	+	I	I	I	I	+
D-Mannose	+	+	(+)	+	+	+	I	+	(+)
D-Mannitol	+	+	Ι	+	+	+	(+)	+	I
Aesculin	+	+	+	I	+	+	+	+	+

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Characteristic	1	2	3	4	ŝ	9	7	8	6
Gluconate	+	+	I	+	+	+	+	+	+
Adipate	Ι	(+)	I	(+)	I	I	I	I	I
Phenylacetate	I	+	I	Ι	I	I	I	I	I
Polar lipids†	DPG, PI, L	DPG, PE, PI, PL, L	DPG, PI, PIM, PL, L	DPG, PG, GL	DPG, PG, PI, PL	DPG, PI, GL	DPG, PC, PG, PE, PI	DPG, PE, PI	DPG, PG, PI, PAGL, PL, L
DNA G+C content (mol%)	72.5	74*	69.7	73.4	70.7	72	70.8	70.5	71.6
Data from Lee (2006).									

†DPG, diphosphatidylglycerol; PC, phosphatidylcholine; PE, phosphatidylethanolamine; PG, phosphatidylglycerol; PI, phosphatidylinositol; PIM, phosphatidylinositol mannoside; PL, unknown phospholipid; PAGL, phosphoaminoglycolipid; GL, unidentified glycolipid; L, unknown lipid

TSA and R2A agar medium are shown in Table S1. The DNA G + C content of strain IP6SC6^T was 72.5 mol%. The major fatty acids and polar lipids of P. jejuensis NRRL B-24460^T were similar to those previously reported (Lee, 2006). The differences in the proportion of fatty acids and slight differences in the types of polar lipids may be due to the different experimental conditions used.

The chemotaxonomic characteristics of IP6SC6^T, such as meso-diaminopimelic acid as the diagnostic diamino acid of the cell-wall peptidoglycan, MK-8(H_4) as the predominant menaguinone, and diphosphatidylglycerol and phosphatidylinositol as major polar lipids, were consistent with those of members of the genus Phycicoccus. The DNA G + C content of strain IP6SC6^T was also within the range of values (69.7-74.0 mol%) reported for recognized species of the genus Phycicoccus (Lee, 2013). The results of the phylogenetic analysis based on 16S rRNA gene sequences suggested that strain IP6SC6^T belonged to the genus Phycicoccus. However, the relatively low levels of sequence similarity (<97 %) between strain IP6SC6^T and recognized species of the genus Phycicoccus indicated that strain IP6SC6^T represents a novel species. Strain IP6SC6^T showed a range of characteristics that differentiated it from other species of the genus *Phycicoccus* (Table 1), including differences either in cultural, physiological and biochemical characteristics, such as cell morphology, temperature and pH range for growth, oxidase activity, hydrolysis of starch, nitrate reduction, carbon source assimilation and enzyme production, or in some chemotaxonomic characteristics, such as the polar lipid pattern and major fatty acid components. Strain IP6SC6^T contained diphosphatidylglycerol, the diagnostic phospholipid of the genus Phycicoccus (Zhang et al., 2011), and phosphatidylinositol, which is found in all members of the genus Phycicoccus except P. bigeumensis MSL-03^T (Dastager et al., 2008); but the absence of, among others, phosphatidylglycerol and phosphatidylethanolamine clearly distinguished it from the other members of the genus *Phycicoccus* (Table 1). The most abundant cellular fatty acids in strain IP6SC6^T were iso- $C_{15:0}$, anteiso- $C_{15:0}$, anteiso- $C_{17:0}$ and iso- $C_{16:0}$ (Table S1). The presence of iso- $C_{15:0}$ and iso- $C_{16:0}$ as the major fatty acids in strain IP6SC6^T and *P. jejuensis* NRRL B-24460^T was a common characteristic shared by all members of the genus Phycicoccus (Table S2) and was also in agreement with the emended description of the genus Phycicoccus given by Zhang et al. (2011). Among the major fatty acids in strain IP6SC6^T, anteiso-C_{15:0} was also detected as a main component in P. ginsenosidimutans KCTC 19419^T (Wang *et al.*, 2011), whereas antesio- $C_{17,0}$ was only detected as a main component in the novel strain, representing a key characteristic differentiating it from all members of the genus Phycicoccus.

Despite the high 16S rRNA gene sequence similarity between strain $IP6SC6^T$ and several species of the genus Tetrasphaera, the new isolate is not affiliated to the genus Tetrasphaera because it is not only grouped into a different clade in all three tree-making methods, but also shows an

Table 1. cont.

absence of the following diagnostic characteristics for species of the genus *Tetrasphaera*. (1) Cells of *Tetrasphaera* species are cocci, occurring singly or in pairs but predominantly as tetrads or clusters, or exhibit a morphological change from rod to coccus shapes (Ishikawa & Yokota, 2006). (2) Members of the genus *Tetrasphaera* grow very slowly and utilize only a limited number of substrates (Maszenan *et al.*, 2000; Ishikawa & Yokota, 2006). (3) *Tetrasphaera duodecadis* IAM 14868^T indispensably requires vitamin B₁₂ for robust growth (Lochhead, 1958; Ishikawa & Yokota, 2006) and isoprenoid quinones cannot be detected in *Tetrasphaera veronensis* Ver1^T (McKenzie *et al.* 2006).

In conclusion, on the basis of phylogenetic analyses, and phenotypic and chemotaxonomic characteristics, strain IP6SC6^T represents a novel species of the genus *Phycicoccus*, for which the name *Phycicoccus endophyticus* sp. nov. is proposed.

Description of Phycicoccus endophyticus sp. nov.

Phycicoccus endophyticus (en.do.phy'ti.cus. Gr. pref. *endo* within; Gr. n. *phyton* plant; L. masc. suff. *-icus* adjectival suffix used with the sense of belonging to; N.L. masc. adj. *endophyticus* within plant, endophytic, pertaining to the isolation from plant tissues).

Cells are Gram-stain-positive, aerobic, non-spore-forming, non-motile rods (0.5–0.8 \times 1.2–2.0 μ m). Substrate and aerial mycelia are not observed, and no diffusible pigment is produced on any of the media tested. Colonies on TSA are circular, smooth, opaque and greyish yellow. Displays good growth on TSA, R2A agar, ISP 2 agar, ISP 3 agar, Bennett's agar and yeast-starch agar, poor growth on ISP 5 agar and ISP 7 agar, and no growth on ISP 4 agar or nutrient agar. Growth occurs at 20-42 °C (optimum, 37 °C), at pH 6.0-9.0 (optimum, pH 7.0) and in the presence of 0-8 % (w/v) NaCl (optimum, 0-2 %). Cells are positive for catalase and weakly positive for oxidase. Tweens 20, 40 and 80, casein and gelatin are hydrolysed, but not starch. Nitrate reduction and H₂S production are negative. Urease and arginine dihydrolase activities are absent. Glucose fermentation does not occur. Milk peptonization and coagulation and aesculin degradation are observed. Assimilates D-glucose, D-mannose, D-mannitol, N-acetyl-D-glucosamine, maltose, gluconate and malate. Does not assimilate D-arabinose, caprate, adipate, citrate or phenylacetate (API 20NE). Positive for esterase (C4), esterase lipase (C8), leucine arylamidase, valine arylamidase (weakly), cysteine arylamidase, acid phosphatase, naphthol-AS-BI-phosphohydrolase, α -glucosidase and β -glucosidase, but negative for alkaline phosphatase, lipase (C14), trypsin, α -chymotrypsin, α -galactosidase, β -galactosidase, β -glucuronidase, N-acetyl- β -glucosaminidase, α -mannosidase and α -fucosidase (API ZYM). Acid is produced from D-arabinose (weakly), D-xylose, D-galactose, D-glucose, D-fructose, D-mannose, mannitol, methyl α-D-glucoside, N-acetylglucosamine (weakly), amygdalin (weakly), arbutin, aesculin,

salicin, cellobiose, maltose, lactose, sucrose, trehalose, xylitol, gentiobiose, turanose, D-lyxose, D-tagatose and D-arabitol (API 50CH). The cell-wall peptidoglycan contains *meso*-diaminopimelic acid as the diagnostic diamino acid. The whole-cell-wall sugars are galactose and glucose. The predominant menaquinone is MK-8(H₄). The polar lipids comprise diphosphatidylglycerol, phosphatidylinositol and an unknown lipid. The major fatty acids (>10 %) are iso- $C_{15:0}$, anteiso- $C_{17:0}$ and iso- $C_{16:0}$.

The type strain, IP6SC6^T (=DSM 100020^{T} =CGMCC 4.7300^T), was isolated from surface-sterilized bark of *Bruguiera gymnorhiza* collected from Zhanjiang Mangrove Forest National Nature Reserve in Guangdong, China. The G+C content of the genomic DNA of the type strain is 72.5 mol%.

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References

Ara, I. & Kudo, T. (2007). *Krasilnikovia* gen. nov., a new member of the family *Micromonosporaceae* and description of *Krasilnikovia cinnamonea* sp. nov. *Actinomycetologica* **21**, 1–10.

Cappuccino, J. G. & Sherman, N. (2002). *Microbiology: a Laboratory Manual*, 6th edn. San Francisco: Benjamin Cummings Pearson Education.

Collins, M. D., Pirouz, T., Goodfellow, M. & Minnikin, D. E. (1977). Distribution of menaquinones in actinomycetes and corynebacteria. *J Gen Microbiol* **100**, 221–230.

Dastager, S. G., Lee, J.-C., Ju, Y.-J., Park, D.-J. & Kim, C.-J. (2008). *Phycicoccus bigeumensis* sp. nov., a mesophilic actinobacterium isolated from Bigeum Island, Korea. *Int J Syst Evol Microbiol* 58, 2425–2428.

Felsenstein, J. (1981). Evolutionary trees from DNA sequences: a maximum likelihood approach. J Mol Evol 17, 368–376.

Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **39**, 783–791.

Fitch, W. M. (1971). Toward defining the course of evolution: minimum change for a specific tree topology. *Syst Zool* 20, 406–416.

Gonzalez, C., Gutierrez, C. & Ramirez, C. (1978). Halobacterium vallismortis sp. nov. An amylolytic and carbohydrate-metabolizing, extremely halophilic bacterium. *Can J Microbiol* 24, 710–715.

Gordon, R. E. & Smith, M. M. (1955). Proposed group of characters for the separation of *Streptomyces* and *Nocardia*. J Bacteriol 69, 147–150.

Guo, L., Tuo, L., Habden, X., Zhang, Y., Liu, J., Jiang, Z., Liu, S., Dilbar, T. & Sun, C. (2015). *Allosalinactinospora lopnorensis* gen. nov., sp. nov., a new member of the family *Nocardiopsaceae* isolated from soil. *Int J Syst Evol Microbiol* 65, 206–213.

Ishikawa, T. & Yokota, A. (2006). Reclassification of Arthrobacter duodecadis Lochhead 1958 as Tetrasphaera duodecadis comb. nov.

and emended description of the genus *Tetrasphaera*. Int J Syst Evol Microbiol 56, 1369–1373.

Kelly, K. L. (1964). Inter-Society Color Council – National Bureau of Standards Color Name Charts illustrated with Centroid Colors. Washington, DC: US Government Printing Office.

Kim, O.-S., Cho, Y.-J., Lee, K., Yoon, S.-H., Kim, M., Na, H., Park, S.-C., Jeon, Y. S., Lee, J.-H. & other authors (2012). Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylotypes that represent uncultured species. *Int J Syst Evol Microbiol* 62, 716–721.

Kimura, M. (1983). The Neutral Theory of Molecular Evolution. Cambridge: Cambridge University Press.

Lee, S. D. (2006). *Phycicoccus jejuensis* gen. nov., sp. nov., an actinomycete isolated from seaweed. *Int J Syst Evol Microbiol* 56, 2369–2373.

Lee, S. D. (2013). *Phycicoccus badiiscoriae* sp. nov., a novel actinomycete isolated from scoria. *Int J Syst Evol Microbiol* 63, 989–994.

Li, W.-J., Xu, P., Schumann, P., Zhang, Y.-Q., Pukall, R., Xu, L.-H., Stackebrandt, E. & Jiang, C.-L. (2007). *Georgenia ruanii* sp. nov., a novel actinobacterium isolated from forest soil in Yunnan (China), and emended description of the genus *Georgenia*. Int J Syst Evol Microbiol 57, 1424–1428.

Lochhead, A. G. (1958). Two new species of *Arthrobacter* requiring respectively vitamin B_{12} and the terregens factor. *Arch Mikrobiol* **31**, 163–170.

Magee, C. M., Rodeheaver, G., Edgerton, M. T. & Edlich, R. F. (1975). A more reliable gram staining technic for diagnosis of surgical infections. *Am J Surg* 130, 341–346.

Marmur, J. (1961). A procedure for the isolation of deoxyribonucleic acid from microorganisms. *J Mol Biol* **3**, 208–218.

Maszenan, A. M., Seviour, R. J., Patel, B. K. C., Schumann, P., Burghardt, J., Tokiwa, Y. & Stratton, H. M. (2000). Three isolates of novel polyphosphate-accumulating gram-positive cocci, obtained from activated sludge, belong to a new genus, *Tetrasphaera* gen. nov., and description of two new species, *Tetrasphaera japonica* sp. nov. and *Tetrasphaera australiensis* sp. nov. *Int J Syst Evol Microbiol* **50**, 593–603.

McKenzie, C. M., Seviour, E. M., Schumann, P., Maszenan, A. M., Liu, J.-R., Webb, R. I., Monis, P., Saint, C. P., Steiner, U. & Seviour, R. J. (2006). Isolates of *'Candidatus Nostocoida limicola'* Blackall *et al.* 2000 should be described as three novel species of the genus *Tetrasphaera*, as *Tetrasphaera jenkinsii* sp. nov., *Tetrasphaera vanveenii* sp. nov. and *Tetrasphaera veronensis* sp. nov. Int J Syst Evol *Microbiol* 56, 2279–2290.

Mesbah, M., Premachandran, U. & Whitman, W. B. (1989). Precise measurement of the G+C content of deoxyribonucleic acid by high-performance liquid chromatography. *Int J Syst Bacteriol* **39**, 159–167.

Minnikin, D. E., O'Donnell, A. G., Goodfellow, M., Alderson, G., Athalye, M., Schaal, A. & Parlett, J. H. (1984). An integrated procedure for the extraction of bacterial isoprenoid quinones and polar lipids. *J Microbiol Methods* **2**, 233–241.

Qin, S., Li, J., Chen, H.-H., Zhao, G.-Z., Zhu, W.-Y., Jiang, C.-L., Xu, L.-H. & Li, W.-J. (2009). Isolation, diversity, and antimicrobial

activity of rare actinobacteria from medicinal plants of tropical rain forests in Xishuangbanna, China. *Appl Environ Microbiol* **75**, 6176–6186.

Saitou, N. & Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4, 406–425.

Sasser, M. (1990). Identification of bacteria by gas chromatography of cellular fatty acids, MIDI Technical Note 101. Newark, DE: MIDI Inc.

Schleifer, K. H. & Kandler, O. (1972). Peptidoglycan types of bacterial cell walls and their taxonomic implications. *Bacteriol Rev* 36, 407–477.

Shirling, E. B. & Gottlieb, D. (1966). Methods for characterization of *Streptomyces* species. *Int J Syst Bacteriol* 16, 313–340.

Singh, H., Won, K., Ngo, H. T. T., Du, J., Kook, M. & Yi, T.-H. (2015). *Phycicoccus soli* sp. nov., isolated from soil. *Int J Syst Evol Microbiol* 65, 2351–2356.

Stackebrandt, E., Rainey, F. A. & Ward-Rainey, N. L. (1997). Proposal for a new hierarchic classification system, *Actinobacteria* classis nov. *Int J Syst Bacteriol* **47**, 479–491.

Staneck, J. L. & Roberts, G. D. (1974). Simplified approach to identification of aerobic actinomycetes by thin-layer chromatography. *Appl Microbiol* **28**, 226–231.

Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013). MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* **30**, 2725–2729.

Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997). The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* **25**, 4876–4882.

Tuo, L., Dong, Y.-P., Habden, X., Liu, J.-M., Guo, L., Liu, X.-F., Chen, L., Jiang, Z.-K., Liu, S.-W. & other authors (2015). *Nocardioides deserti* sp. nov., an actinobacterium isolated from desert soil. *Int J Syst Evol Microbiol* 65, 1604–1610.

Waksman, S. A. (1961). Classification, Identification and Description of Genera and Species. 2, Baltimore: Williams & Wilkins.

Wang, L., An, D.-S., Jin, F.-X., Lee, S.-T., Im, W.-T. & Bae, H.-M. (2011). *Phycicoccus ginsenosidimutans* sp. nov., isolated from soil of a ginseng field. *Int J Syst Evol Microbiol* **61**, 524–528.

Weon, H.-Y., Yoo, S.-H., Kim, B.-Y., Schumann, P., Kroppenstedt, R. M., Hong, S.-K. & Kwon, S.-W. (2008). *Phycicoccus aerophilus* sp. nov., isolated from air. *Int J Syst Evol Microbiol* **58**, 2389–2392.

Xu, P., Li, W.-J., Tang, S.-K., Zhang, Y.-Q., Chen, G.-Z., Chen, H.-H., Xu, L.-H. & Jiang, C.-L. (2005). *Naxibacter alkalitolerans* gen. nov., sp. nov., a novel member of the family '*Oxalobacteraceae*' isolated from China. *Int J Syst Evol Microbiol* 55, 1149–1153.

Yoon, J.-H., Lee, S.-Y., Kang, S.-J. & Oh, T.-K. (2008). Phycicoccus dokdonensis sp. nov., isolated from soil. Int J Syst Evol Microbiol 58, 597–600.

Zhang, J.-Y., Liu, X.-Y. & Liu, S.-J. (2011). *Phycicoccus cremeus* sp. nov., isolated from forest soil, and emended description of the genus *Phycicoccus. Int J Syst Evol Microbiol* 61, 71–75.